



Funded by
the European Union

Bioinformatics Summer School

Long-reads Transcriptomics

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Nottingham, UK

- 1 Library Preparations
- 2 Basecalling

Section 1

Library preparation (lib-prep)

Library preparation strategies for long-reads RNA-seq experiments

What is your goal?



RNA-seq

Main difference with short-reads lib-prep

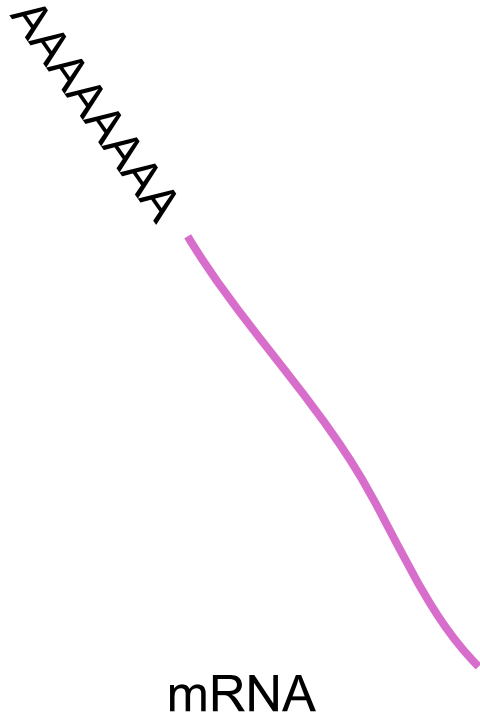


Image: PacBio

RNA-seq

Main difference with short-reads lib-prep

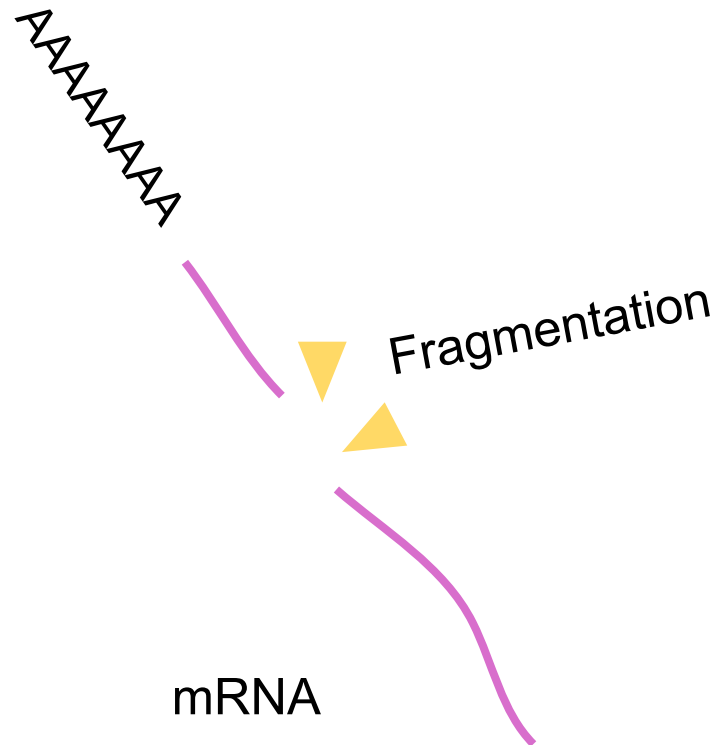


Image: Thermo

RNA-seq

Main difference with short-reads lib-prep

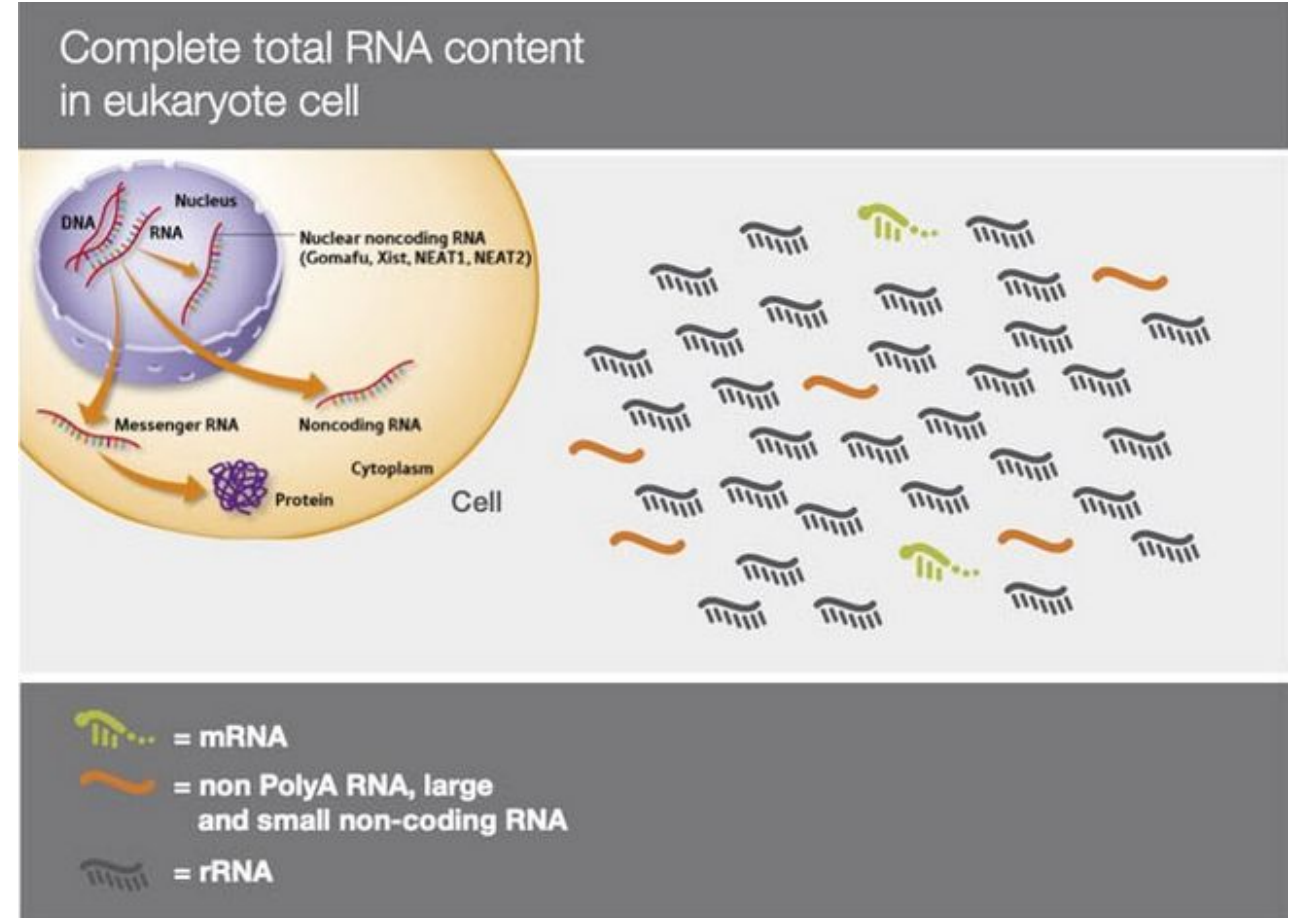
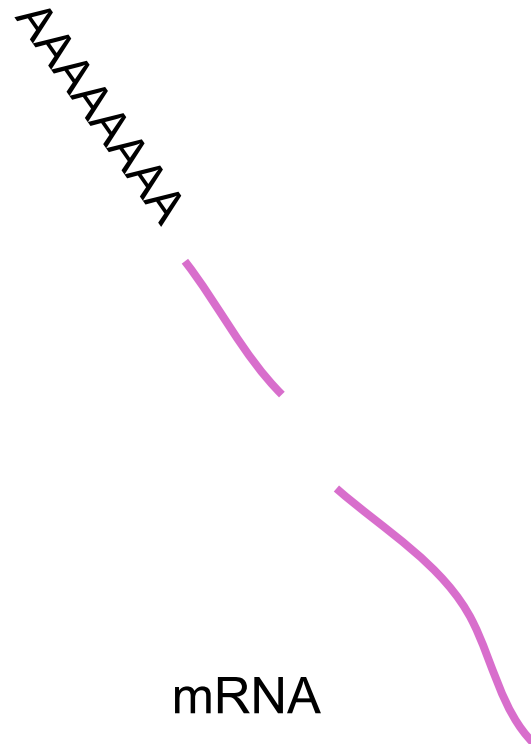


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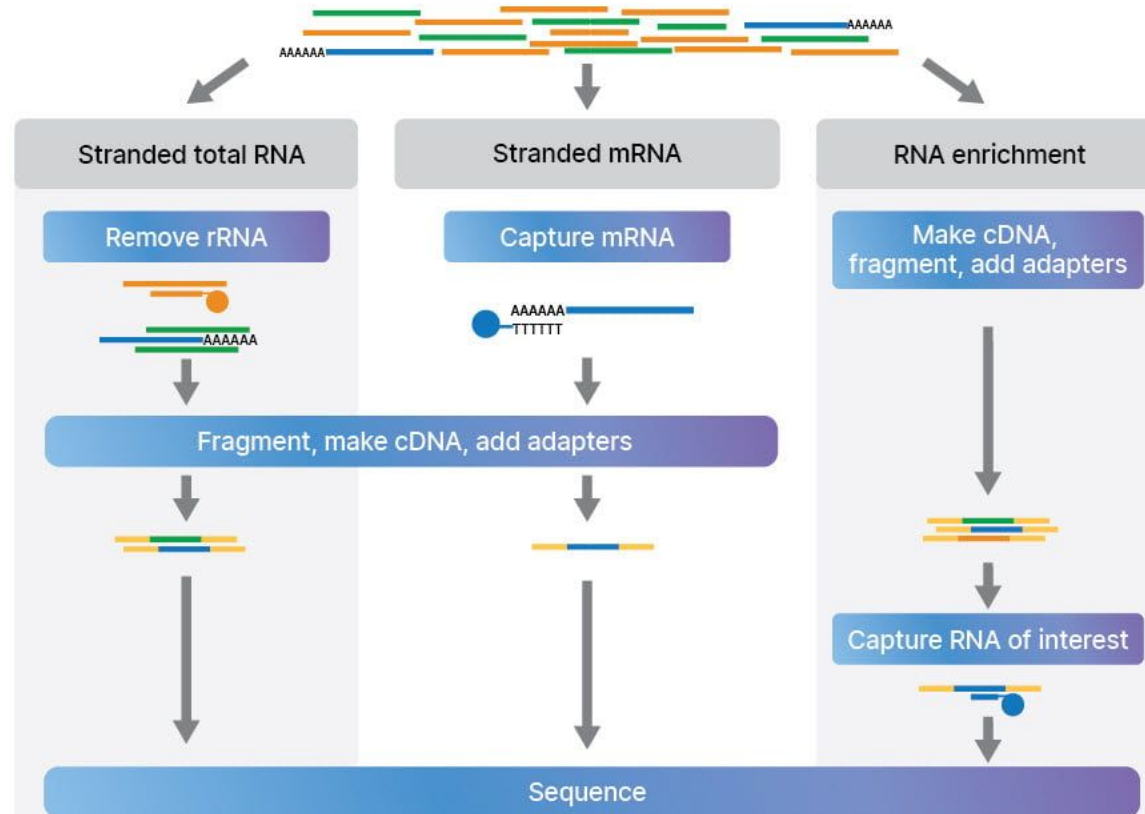
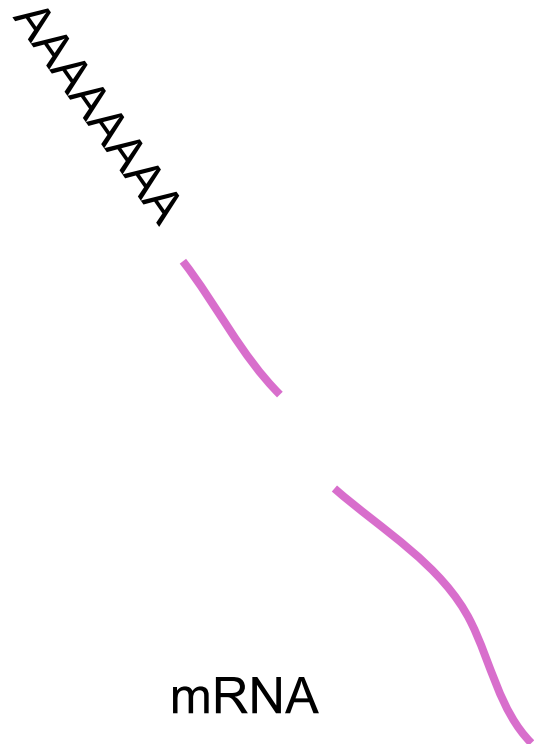


Image: Illumina

Consideration Working with RNA

Garbage in, garbage out



Consideration Working with RNA

Garbage in, garbage out



Consideration Working with RNA

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Consideration Working with RNA

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Consideration Working with RNA

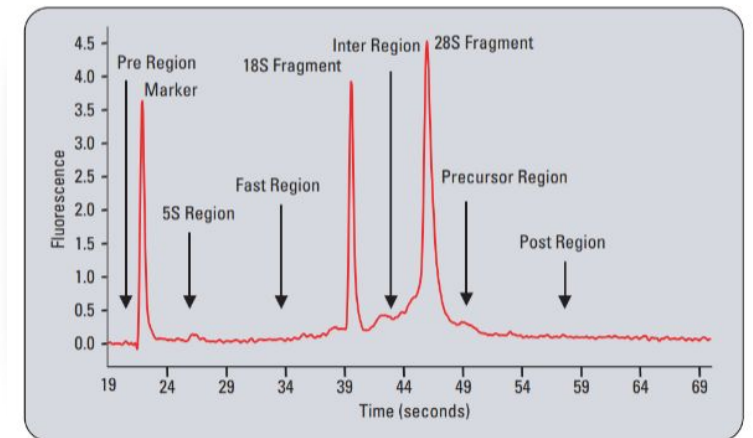
Garbage in, garbage out



RIN < 7

Consideration Working with RNA

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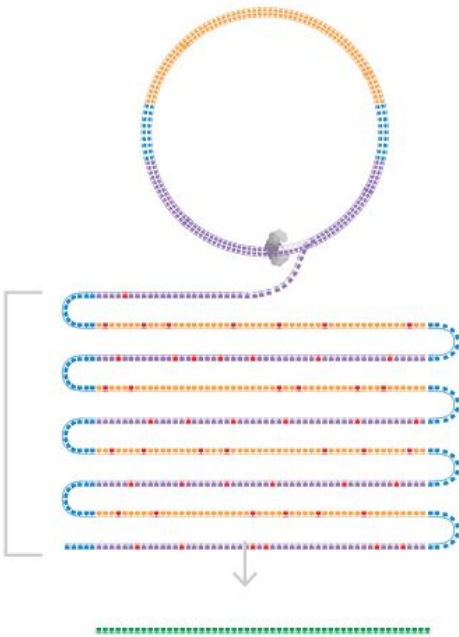


RIN < 7

cDNA
Iso-Seq

PacBio

Description	Full-length transcript isoform sequencing
Kit	SMRTbell prep kit 3.0
Multiplex	up to 12
Preparation time	8 Hours 2 Safe stop point
Input	300 ng Total RNA
Output	1M (Sequel I); 8M (Sequel II); Full-length HiFi transcripts



HiFi read
(99.9% accuracy)

HiFi long reads generated from
PacBio sequencing

Image: PacBio

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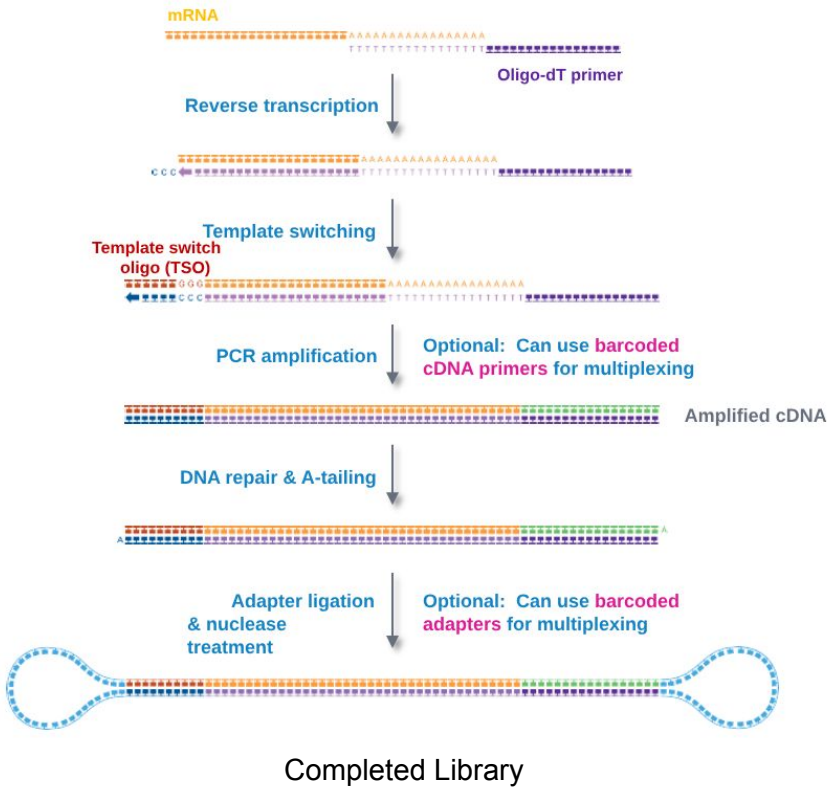
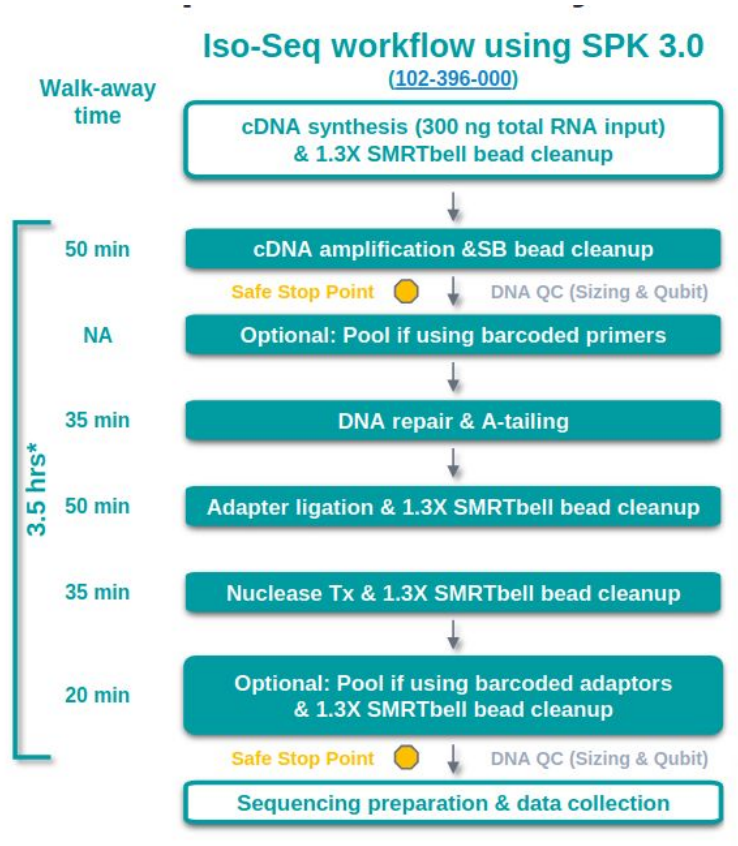


Image: PacBio

cDNA Iso-Seq

PacBio

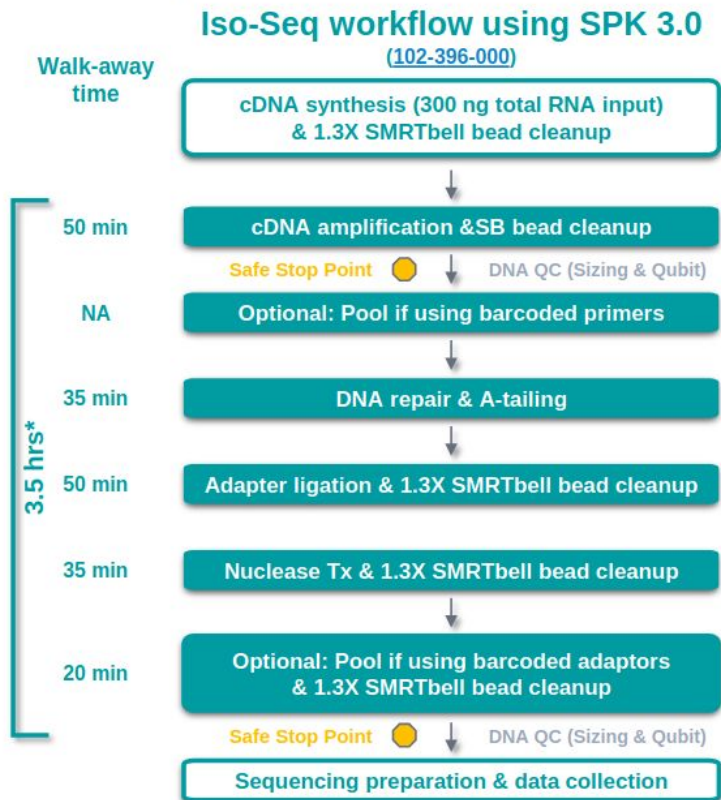


Complete Iso-Seq library preparation workflow for cDNA sequencing

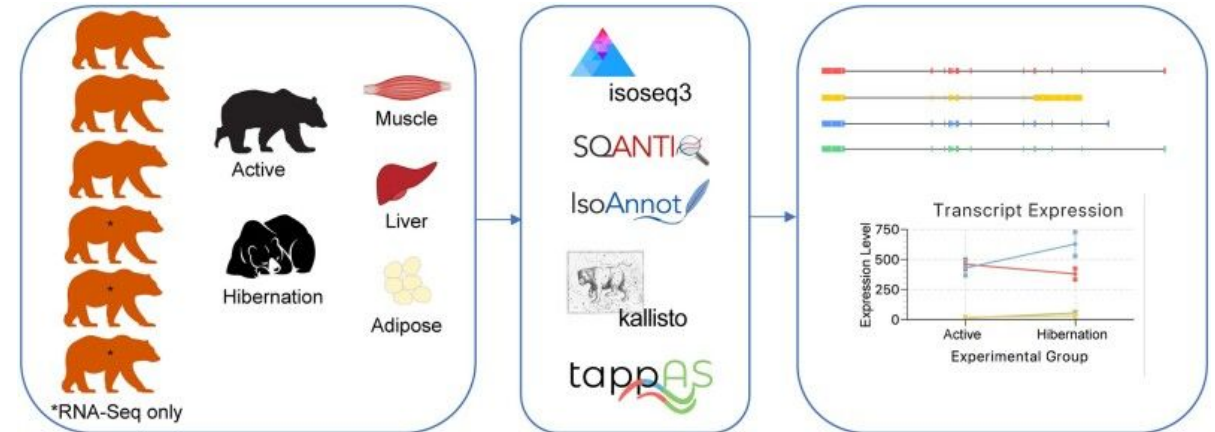
Image: PacBio

cDNA Iso-Seq

PacBio



Complete Iso-Seq library preparation workflow for cDNA sequencing



Adipose is the most dynamic tissue during hibernation with the highest number of genes with differential isoform usage and isoform switching as compared to liver and muscle.

Tseng et al.(2022). Long-read isoform sequencing reveals tissue-specific isoform expression between active and hibernating brown bears (*Ursus arctos*).



Iso-Seq is great



But it can be better

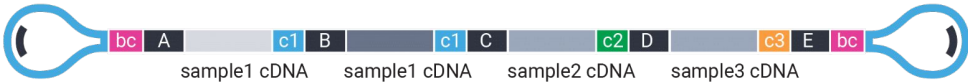
imgflip.com

cDNA

Kinnex (Commercial version of MAS-ISO-seq)

PacBio

Description	Full-length isoform discovery with flexible sample multiplexing
Kit	Kinnex full-length RNA kit
Multiplex	5M × 3-plex (Sequel II); 5M x 6-plex (Vega); 10M × 6-plex (Revio SPRQ)
Preparation time	1.5 days 4 Safe stop point
Input	300 ng Total RNA
Output	15-20M (Sequel II); 20-30M (Vega); 50-60M (Revio SPRQ) Full-length HiFi transcripts (Q40)
Notable Studies	Al Khafaji et al., 2023. Nature Portofolio



Completed Library with cDNA concatenated

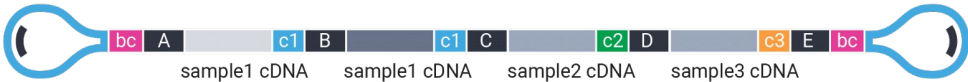
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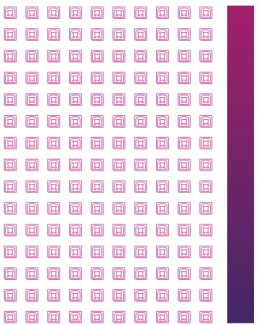
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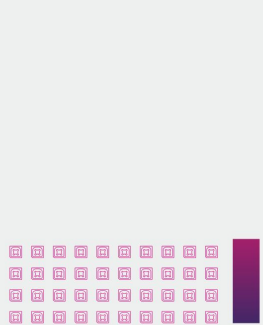
Completed Library with cDNA concatenated

Sequel II system without Kinnex



150 SMRT Cell 8M in 38 weeks

Sequel II system with Kinnex



40 SMRT Cell 8M in 10 weeks

Revio system with Kinnex



11 SMRT Cell 25M in 3 days

Kinnex increase throughput significantly

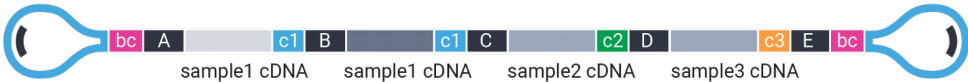
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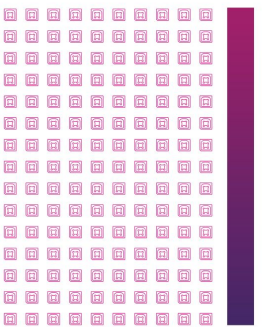
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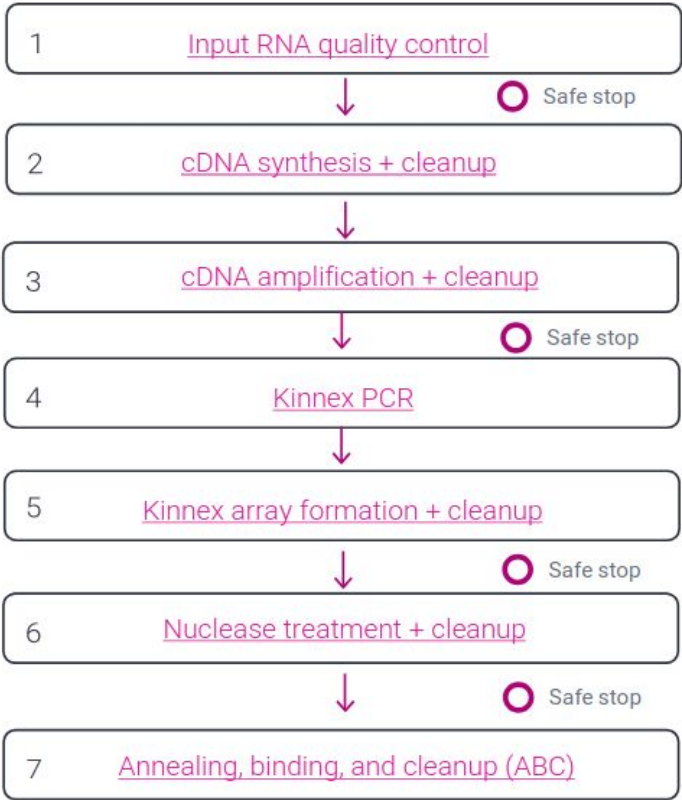
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Image: PacBio

cDNA

Kinnex (Commercial version of MAS-ISO-seq)

PacBio



SMRTbell® library preparation	
Iso-Seq® Express 2.0 Kit	PacBio® 103-071-500
SMRTbell® cleanup beads	PacBio® 102-158-300*
Elution buffer (50 mL)	PacBio® 101-633-500*
Kinnex™ PCR 8-fold kit	PacBio® 103-071-600*
Revio® SPRQ™ polymerase kit or	PacBio® 103-520-100
Vega™ polymerase kit or	PacBio® 103-517-600
Revio® polymerase kit** or	PacBio® 102-817-600
Sequel® II binding kit 3.2**	PacBio® 102-333-300

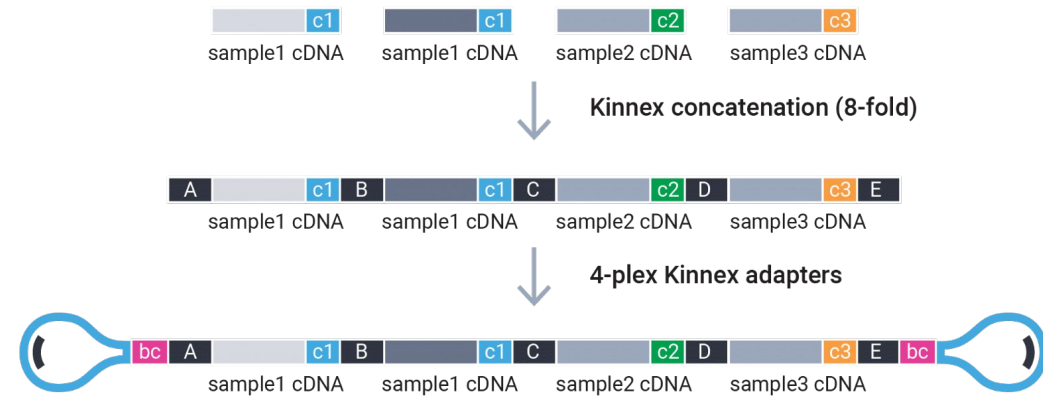
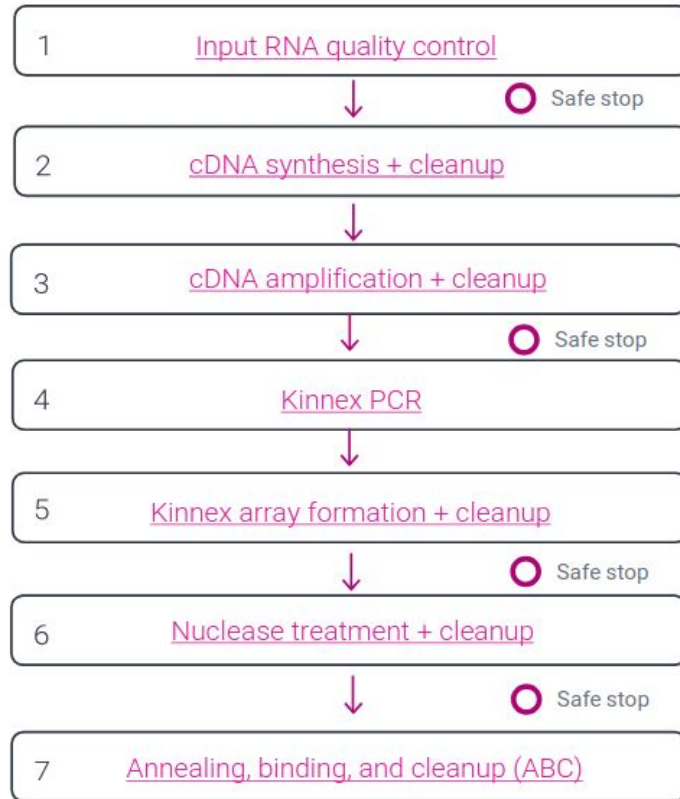
Complete Kinnex library preparation workflow for cDNA sequencing

Image: PacBio

cDNA

Kinnex (Commercial version of MAS-ISO-seq)

PacBio



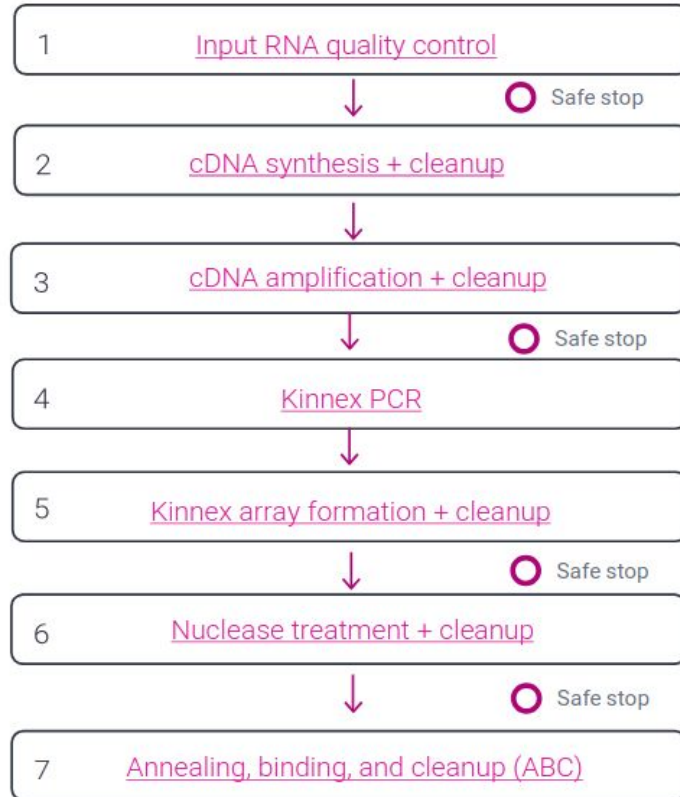
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PacBio



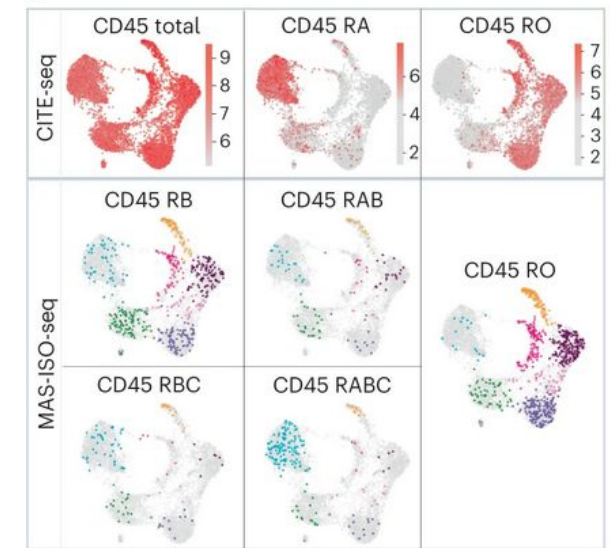
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Image: PacBio

Long-read (MAS-ISO-seq)



c



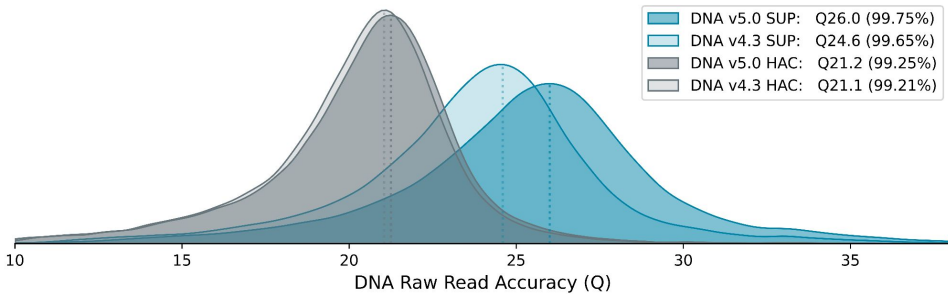
Increasing the throughput >15-fold to nearly 40 million cDNA reads per run. MAS-ISO-seq demonstrated a 12- to 32-fold increase in the discovery of differentially spliced genes.

Al'Khafaji, A.M., Smith, J.T., Garimella, K.V. et al (2024). High-throughput RNA isoform sequencing using programmed cDNA concatenation. <https://doi.org/10.1038/s41587-023-01815-7>

cDNA
cDNA-PCR (R10.4.1)

Nanopore

Description	A sequencing kit optimised for identification and quantification of full-length transcripts with highest output.
Kit	cDNA-PCR Sequencing Kit V14
Multiplex	up to 24-plex (cDNA-PCR Barcoding Kit V14)
Preparation time	225 minutes + PCR
Input	10 ng poly(A)+ RNA or 500 ng total RNA
Output	10-15 million cDNA (MinION); 100 million cDNA (PromethION)



Latest transformer model and R10 pores
deliver higher accuracy

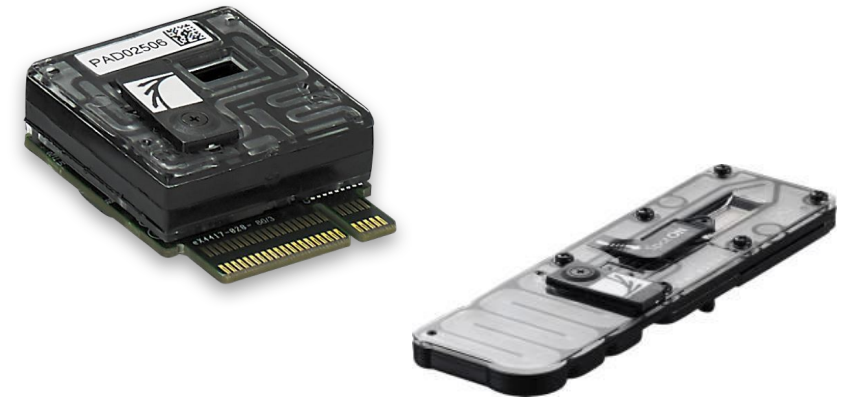
Image: Nanopore

cDNA

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Two types of nanopore device with different sequencing throughput (Left: PromethION, Right: MinION)

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Completed Library

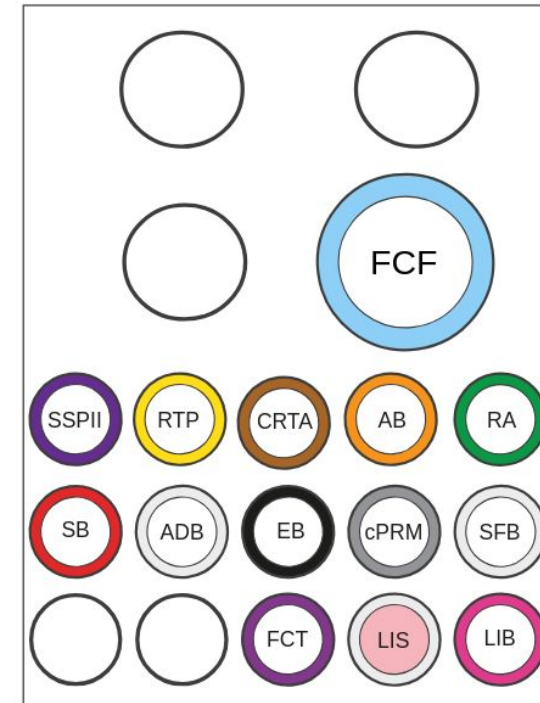
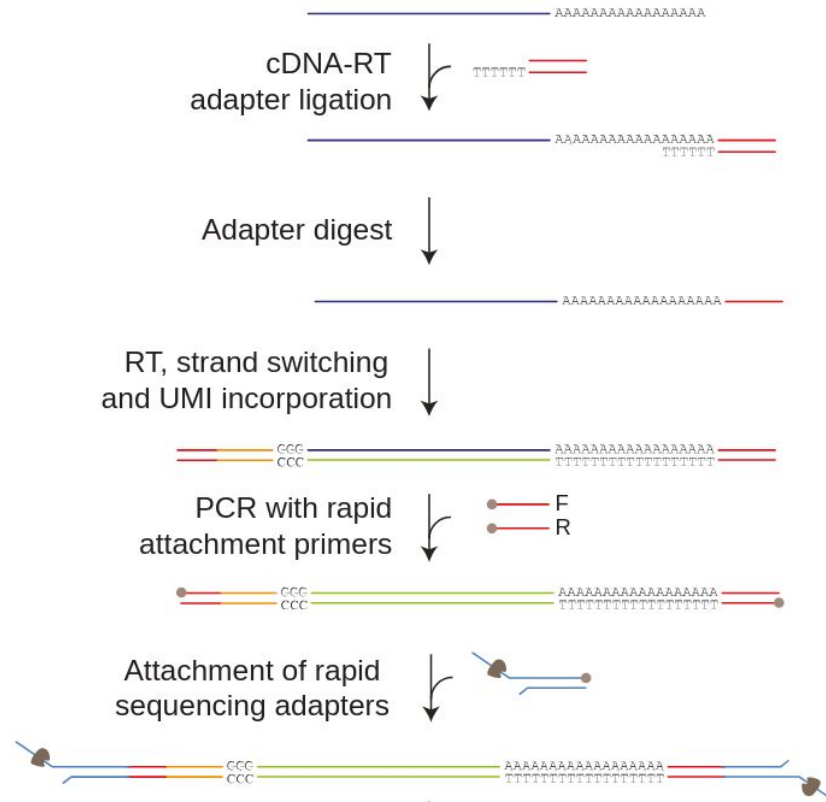
Common Gotchas
Always perform flow cell QC prior lib prep, ensure you have enough pores!

Image: Nanopore

cDNA

cDNA-PCR (R10.4.1)

Nanopore



RTP: RT Primer
 CRTA: cDNA RT Adapter
 AB: Annealing Buffer
 RA: Rapid Adapter
 SB: Sequencing Buffer
 ADB: Adapter Buffer
 SFB: Short Fragment Buffer
 SSPII: Strand Switching Primer II

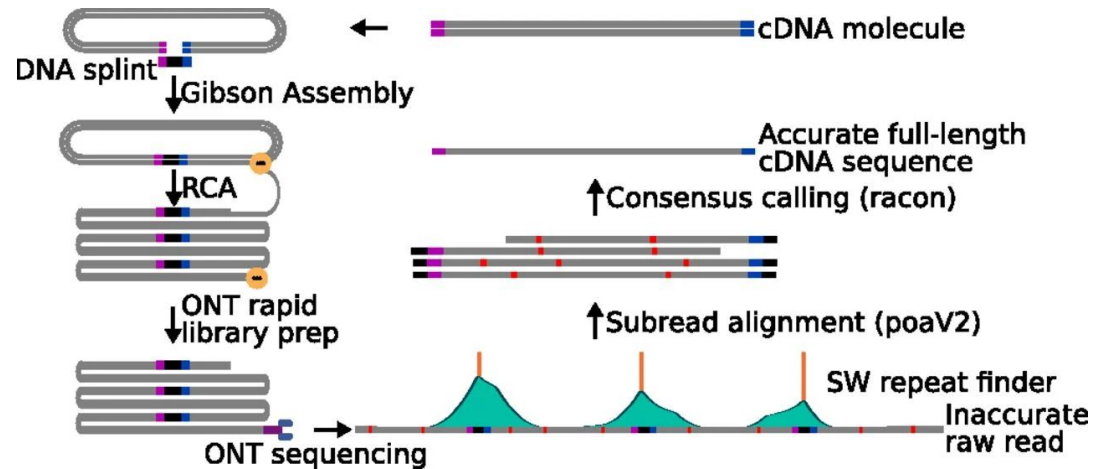
EB: Elution Buffer
 cPRM: cDNA Primer
 LIS: Library Solution
 FCF: Flow Cell Flush
 FCT: Flow Cell Tether

Complete library workflow for cDNA sequencing with cDNA-PCR kit

Image: Nanopore

cDNA

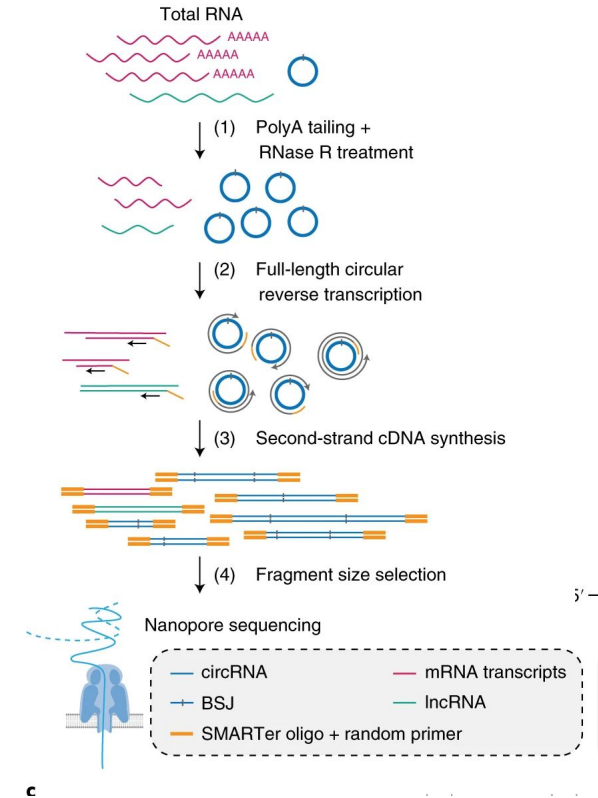
cDNA-PCR (R10.4.1)



Mandalorion was used to analyze the R2C2 reads of the 96 B cells and identified several surface receptor isoforms expressed by 96 distinct single human B cells.



R. Volden, et. al. Improving nanopore read accuracy with the R2C2 method enables the sequencing of highly multiplexed full-length single-cell cDNA. <https://doi.org/10.1073/pnas.1806447115> (2018).



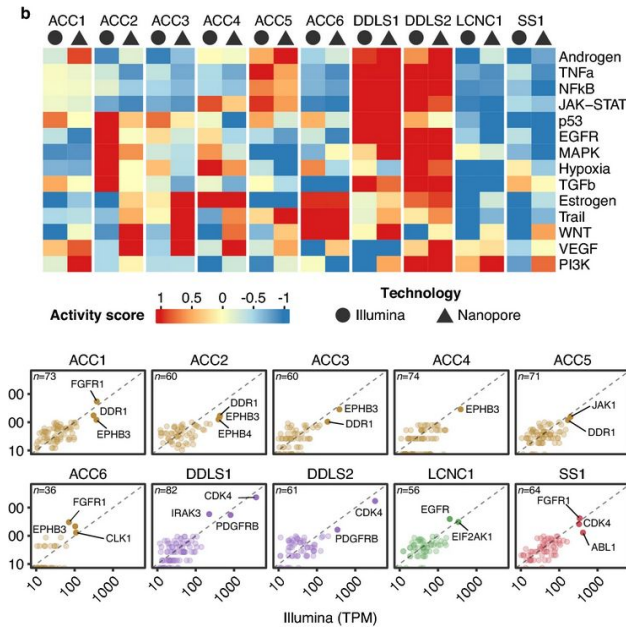
Circular reverse transcription and size selection achieves a 20-fold higher enrichment of circRNAs from total RNA compared to previous methods.

Zhang, J., Hou, L., Zuo, Z. et al. Comprehensive profiling of circular RNAs with nanopore sequencing and CIRI-long. *Nat Biotechnol* 39, 836–845 (2021). <https://doi.org/10.1038/s41587-021-00842-6>

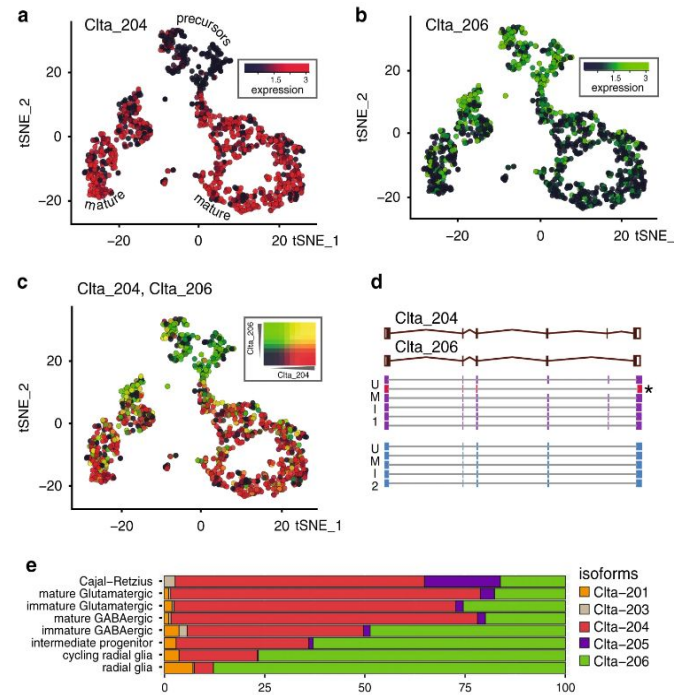
Nanopore
Community

cDNA

cDNA-PCR (R10.4.1)

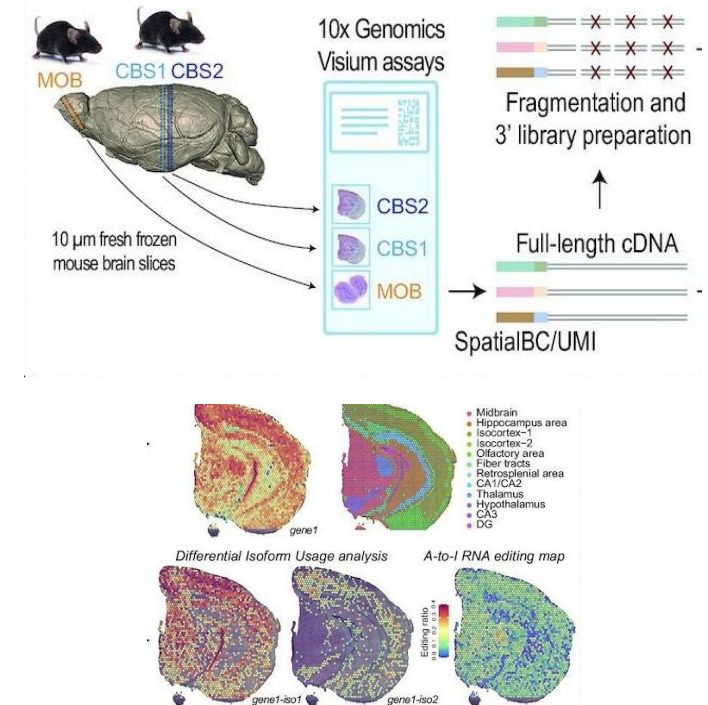


Shallow nanopore RNA-seq enables rapid and biologically meaningful transcriptome profiling of tumors (Mock et al 2023)



Nanopore scRNA-seq reveals transcript isoform diversity (Lebrigand et al 2020).

Nanopore Community



SiT can be used to profile isoform expression and sequence heterogeneity in different areas of the tissue (Lebrigand et al 2023).

mRNA (Native RNA)

Direct RNA sequencing (RNA001, RNA002)

Nanopore

Description	A sequencing kit optimised for sequencing native RNA with improved output and accuracy.
Kit	Direct RNA Sequencing Kit
Multiplex	Under development
Preparation time	135 minutes
Input	300 ng poly(A)+ RNA or 1 ug total RNA
Output	2 GB cDNA (MinION); 30 million cDNA (PromethION)

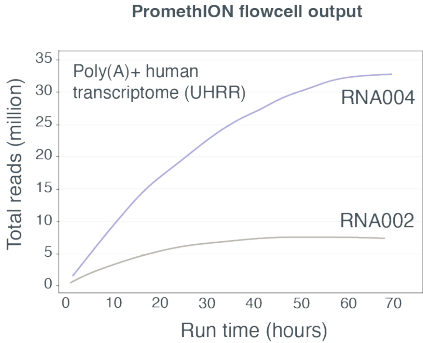
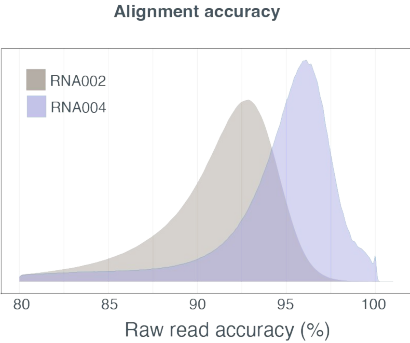
Image: Nanopore

mRNA (Native RNA)

Direct RNA sequencing (RNA004)

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Kit	Direct RNA Sequencing Kit
Multiplex	Under development
Preparation time	135 minutes
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Overall quality and throughput significantly improved in RNA004. Libprep step is the same.

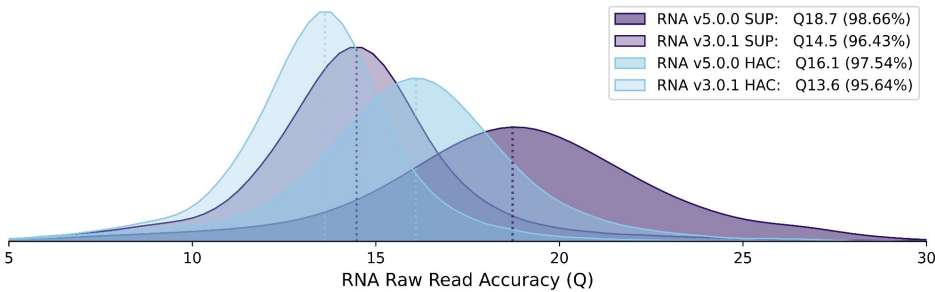
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Nanopore



Latest transformer model and RP4 pores deliver higher accuracy

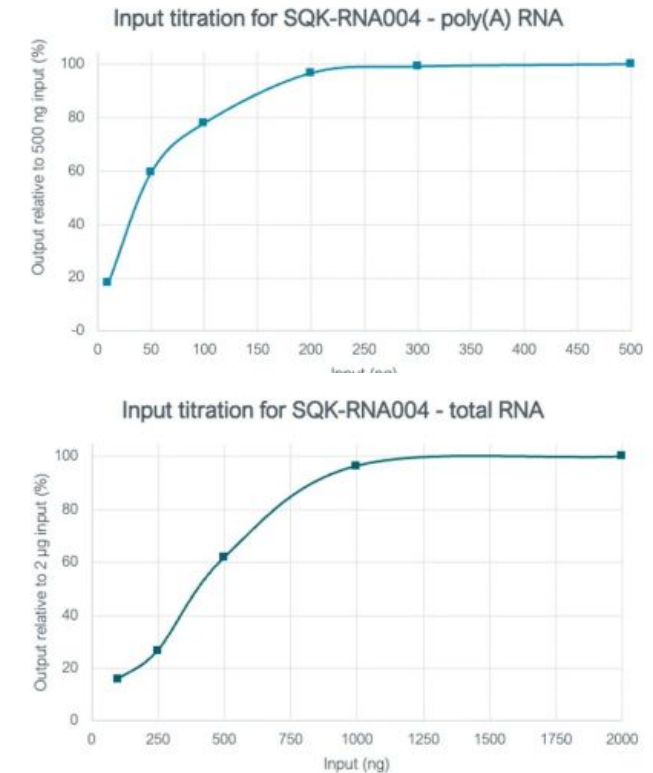
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Direct RNA kit support poly A+ RNA or total RNA

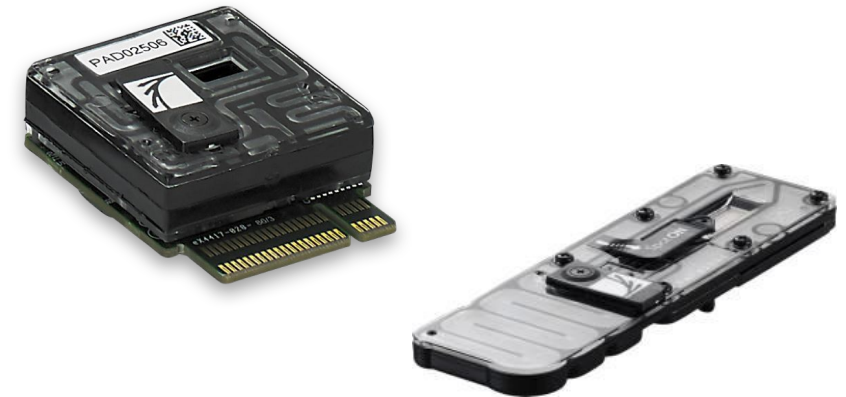
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Two types of nanopore device with different sequencing throughput (Left: PromethION, Right: MinION)

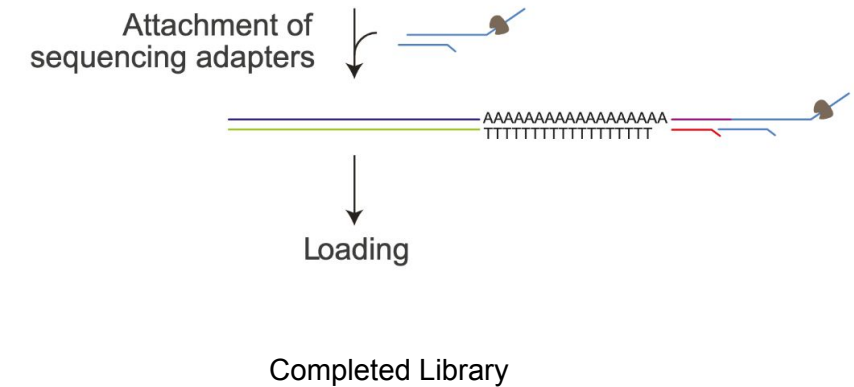
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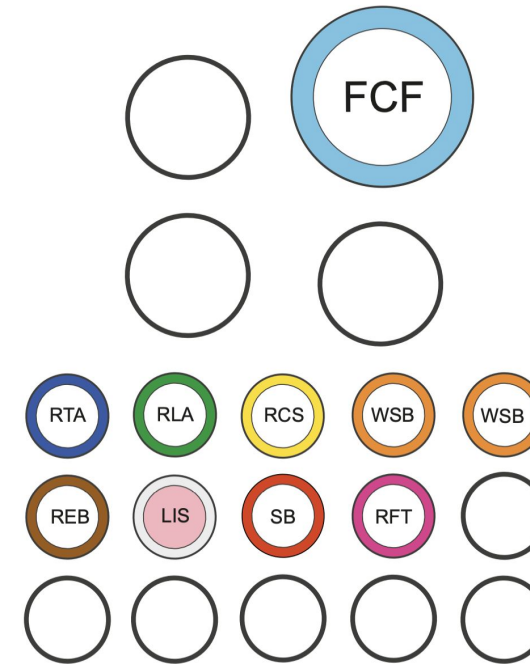
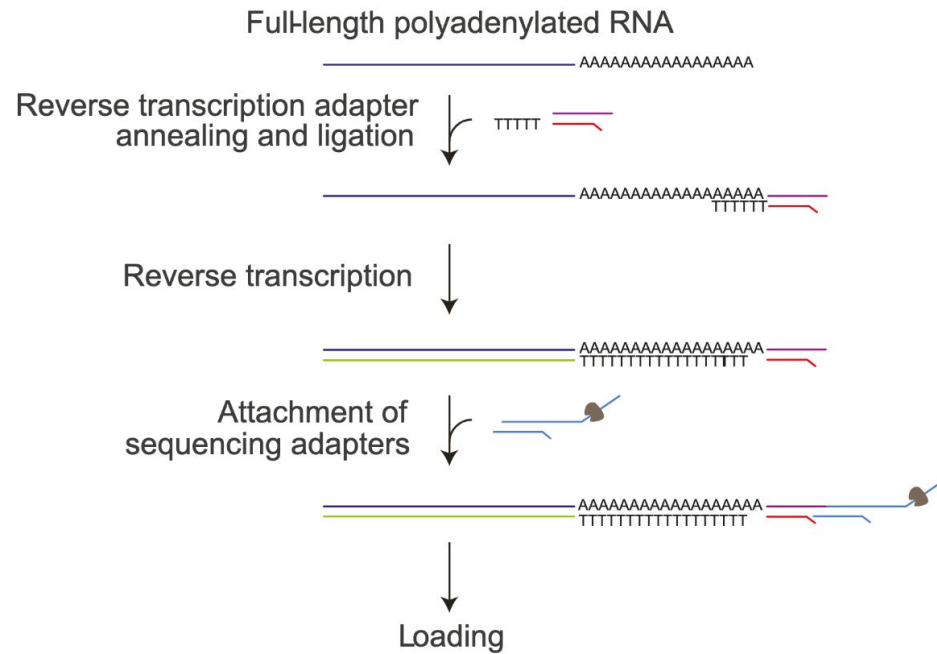
Always perform flow cell QC prior lib prep, ensure you have enough pores!

Image: Nanopore

mRNA (Native RNA)

Direct RNA sequencing (RNA004)

Nanopore



FCF : Flow Cell Flush
RTA: RT Adapter
RLA: RNA Ligation Adapter
RCS: RNA CS
WSB: Wash Buffer

SB : Sequencing Buffer
RFT : RNA Flush Tether
REB : RNA Elution Buffer
LIS : Library Solution

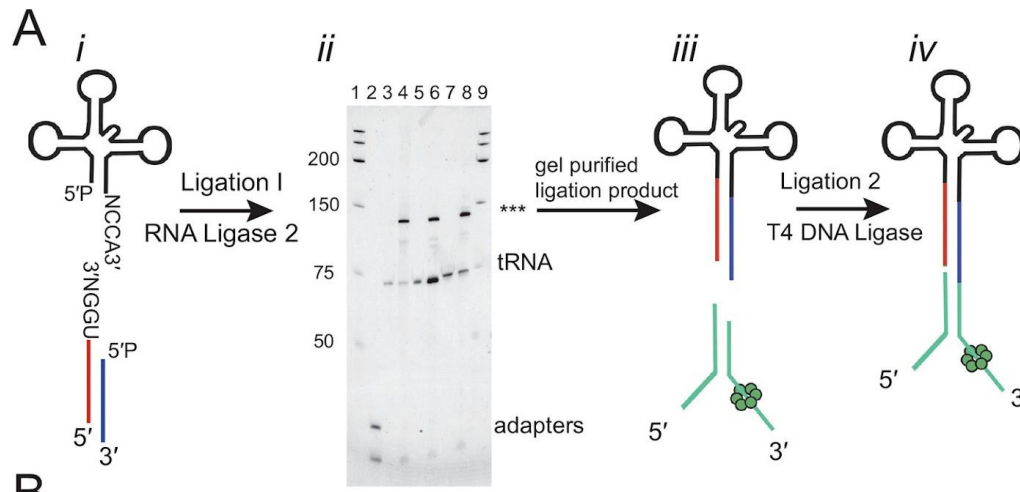
Complete library workflow for Direct RNA sequencing with Direct RNA Sequencing kit

Image: Nanopore

tRNA (Native RNA)

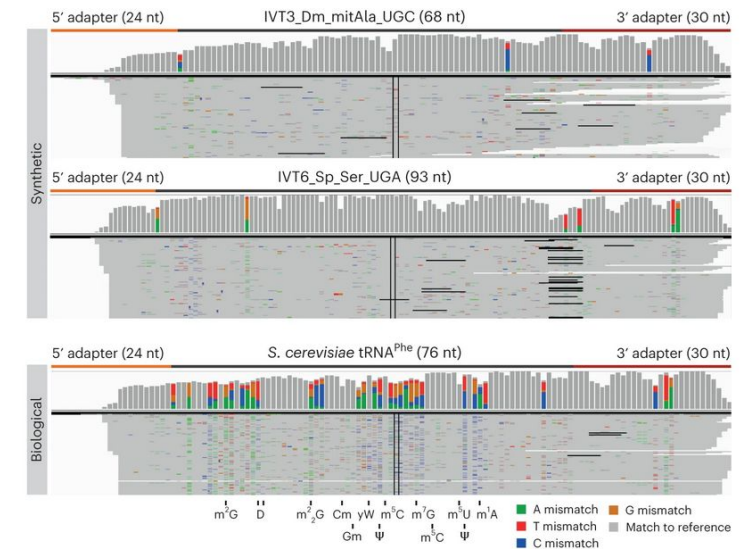
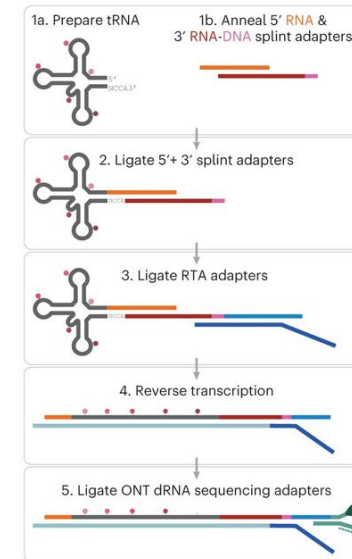
Direct RNA sequencing (RNA004)

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Nanopore sequencing detected all 43 expected isoacceptors in total *E. coli* MRE600 tRNA

Thomas, N. K., Poodari, V. C., Jain, M., Olsen, H. E., Akeson, M., & Abu-Shumays, R. L. (2021). Direct Nanopore Sequencing of Individual Full Length tRNA Strands. *ACS nano*, 15(10), 16642–16653. <https://doi.org/10.1021/acsnano.1c06488>

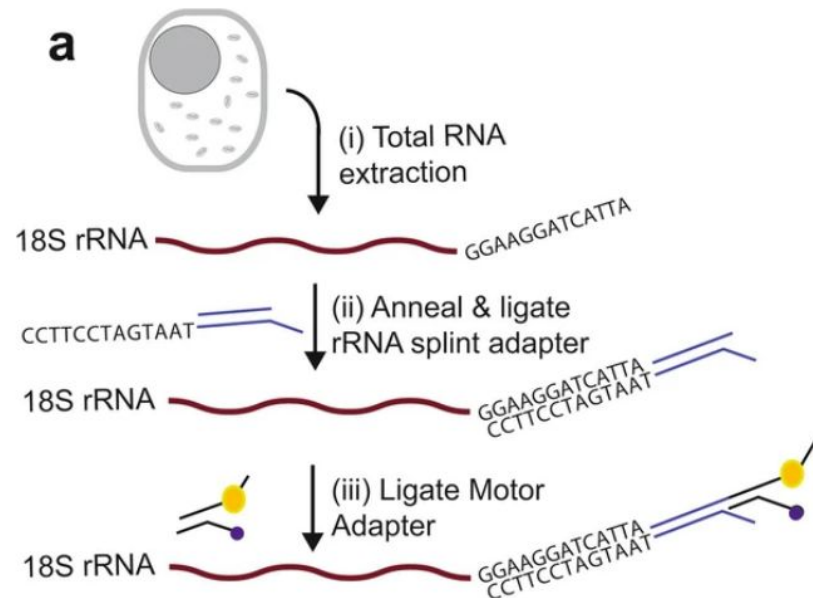


Nano-tRNAseq can efficiently sequence both IVT and native tRNA populations

Lucas, M.C., Pryszcz, L.P., Medina, R. et al. Quantitative analysis of tRNA abundance and modifications by nanopore RNA sequencing. *Nat Biotechnol* 42, 72–86 (2024). <https://doi.org/10.1038/s41587-023-01743-6>

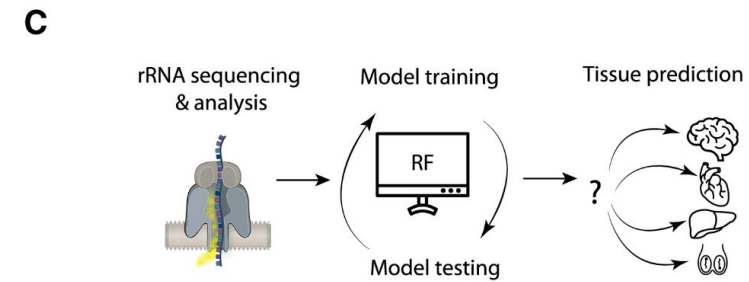
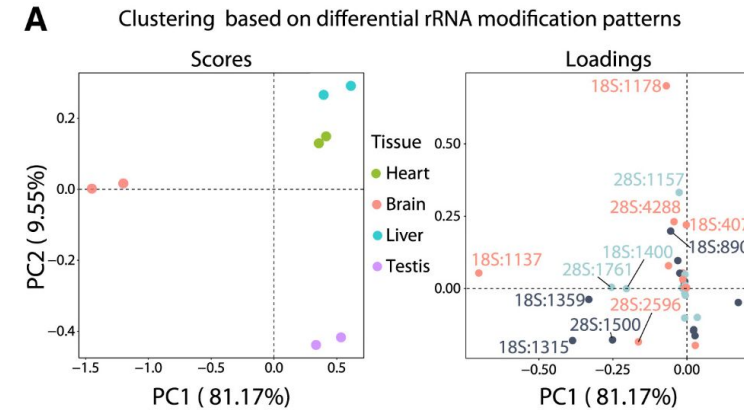
rRNA (Native RNA)

Direct RNA sequencing (RNA004)



Differential rRNA modification patterns can be used to predict tissue types and developmental stages

Jain, M., Olsen, H.E., Akeson, M., Abu-Shumays, R. (2021). Adaptation of Human Ribosomal RNA for Nanopore Sequencing of Canonical and Modified Nucleotides. [Inhttps://doi.org/10.1007/978-1-0716-1374-0_4](https://doi.org/10.1007/978-1-0716-1374-0_4)



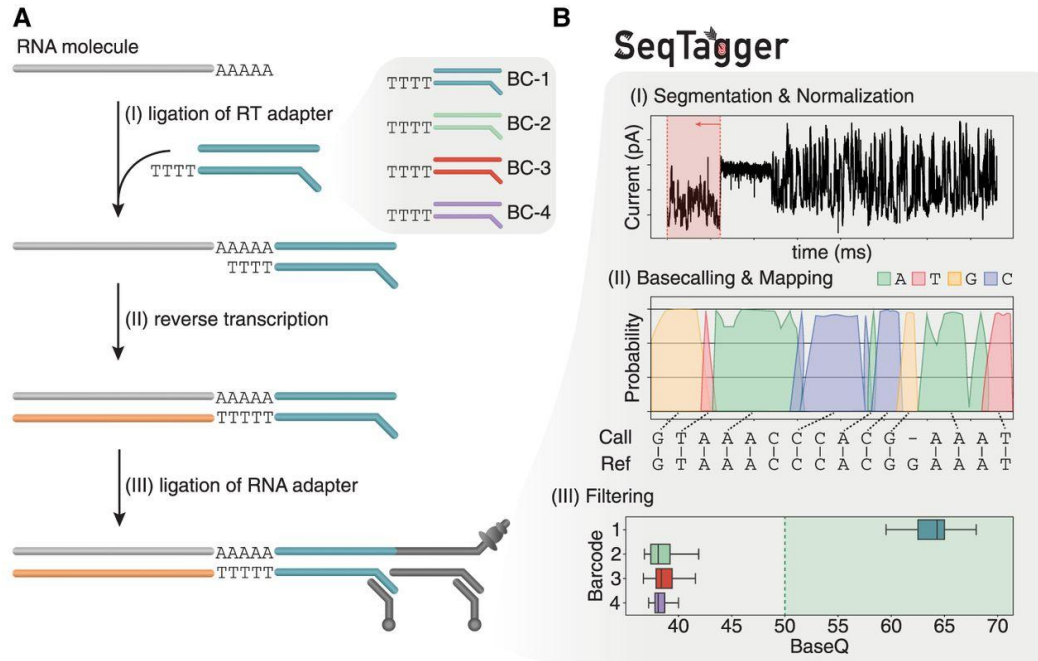
Differential rRNA modification patterns can be used to predict tissue types and developmental stages

Milenkovic, et al. (2025). Epitranscriptomic rRNA fingerprinting reveals tissue-of-origin and tumor-specific signatures. *Molecular cell*, 85(1), 177–190.e7. <https://doi.org/10.1016/j.molcel.2024.11.014>

Nanopore
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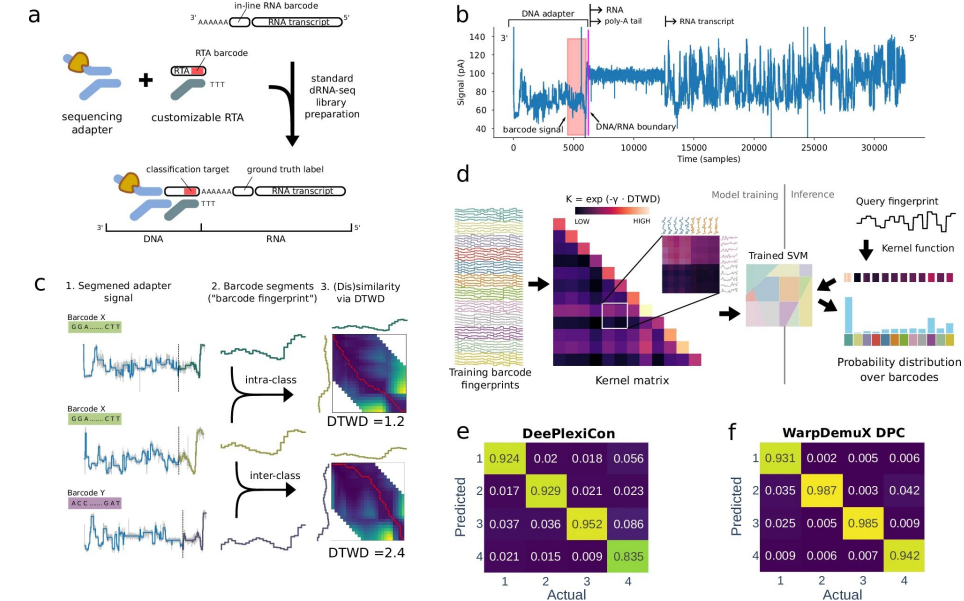
Multiplexing

Direct RNA sequencing (RNA004)



SeqTagger, a rapid and robust method that can demultiplex DRS data sets with 99% precision and 95% recall.

Thomas, N. K., Poodari, V. C., Jain, M., Olsen, H. E., Akeson, M., & Abu-Shumays, R. L. (2021). Direct Nanopore Sequencing of Individual Full Length tRNA Strands. *ACS nano*, 15(10), 16642–16653. <https://doi.org/10.1021/acsnano.1c06488>



Rapid phenotypic profiling of different SARS-CoV-2 viruses through multiplexed sequencing of longitudinal samples on a single flowcell, identifying systematic differences in transcript abundance and poly(A) tail lengths

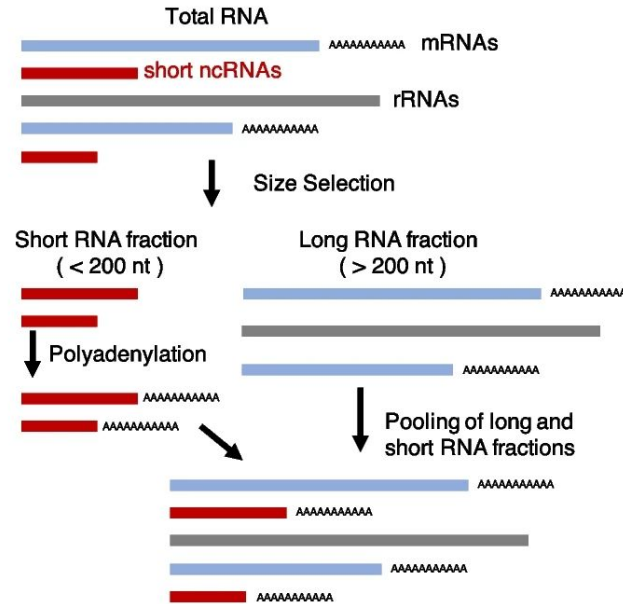
van der Toorn, W., Bohn, P., Liu-Wei, W. et al. Demultiplexing and barcode-specific adaptive sampling for nanopore direct RNA sequencing. *Nat Commun* 16, 3742 (2025). <https://doi.org/10.1038/s41467-025-59102-9>

Improving Representation

Direct RNA sequencing (RNA004)

Nanopore

Community



NERD-seq expands representation of frequently modified non-coding RNAs, such as snoRNAs, snRNAs, scRNAs, srpRNAs, tRNAs, and rRFs.

Saville, L., Wu, L., Habtwold, J. et al. NERD-seq: a novel approach of Nanopore direct RNA sequencing that expands representation of non-coding RNAs. *Genome Biol* 25, 233 (2024).
<https://doi.org/10.1186/s13059-024-03375-8>

Section 1.1

Bonus

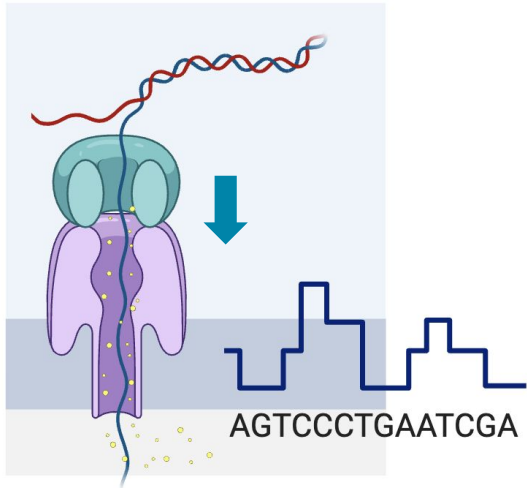
Library preparation strategies for long-reads RNA-seq experiments

Adaptive Sampling

Direct RNA sequencing (RNA004)

Nanopore

Community



Adaptive sampling can be used to enrich or deplete target transcripts.

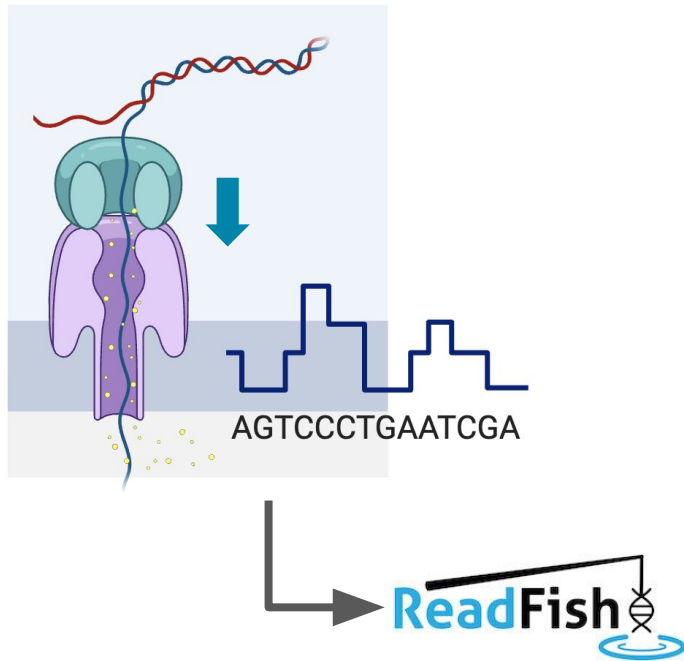
Image: Nanopore

Adaptive Sampling

Direct RNA sequencing (RNA004)

Nanopore

Community



Process data in realtime

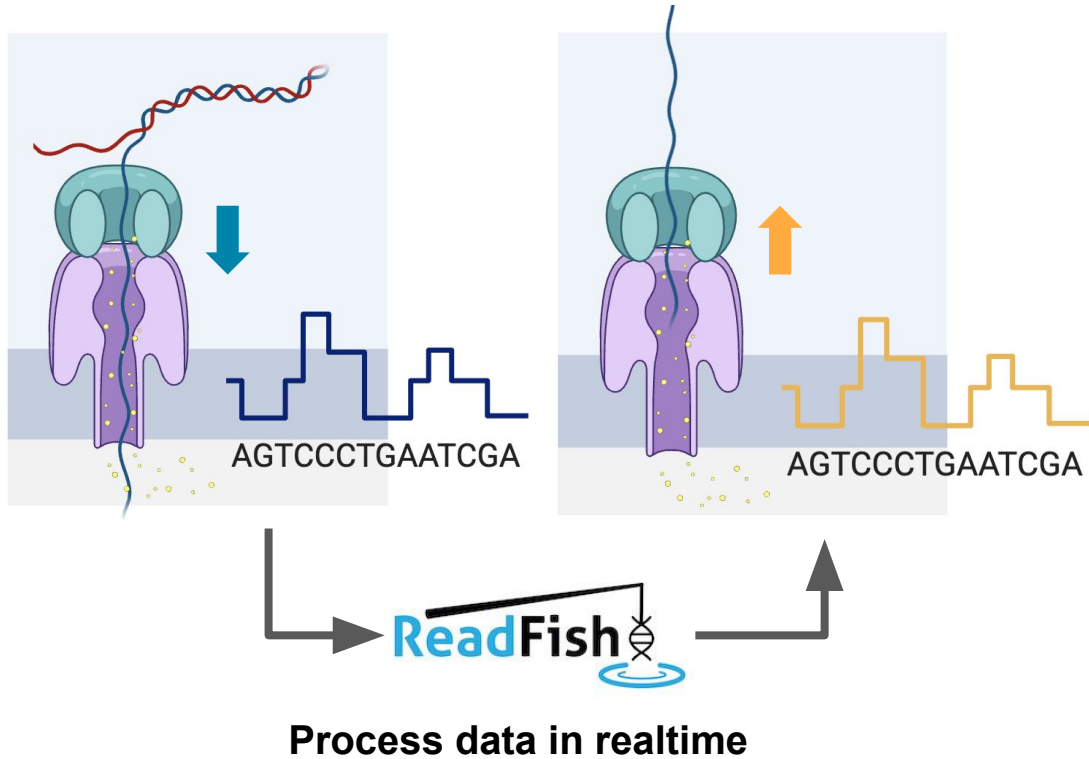
Adaptive sampling can be used to enrich or depletes target transcripts.

Image: Nanopore

Adaptive Sampling

Direct RNA sequencing (RNA004)

Nanopore
Community



Adaptive sampling can be used to enrich or deplete target transcripts.

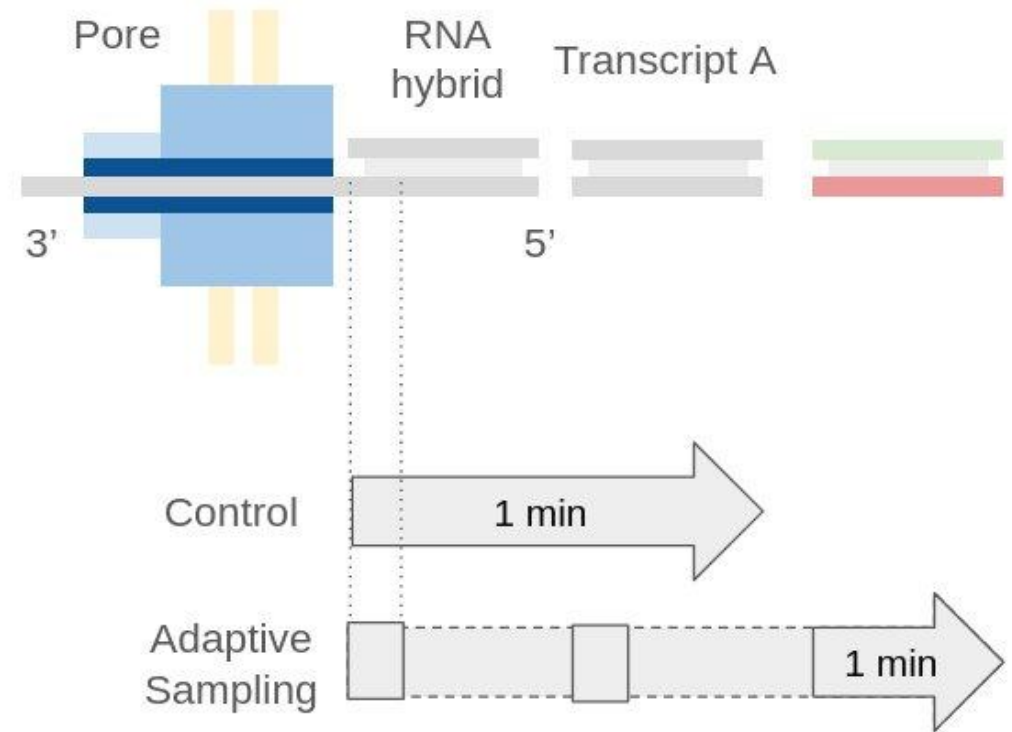
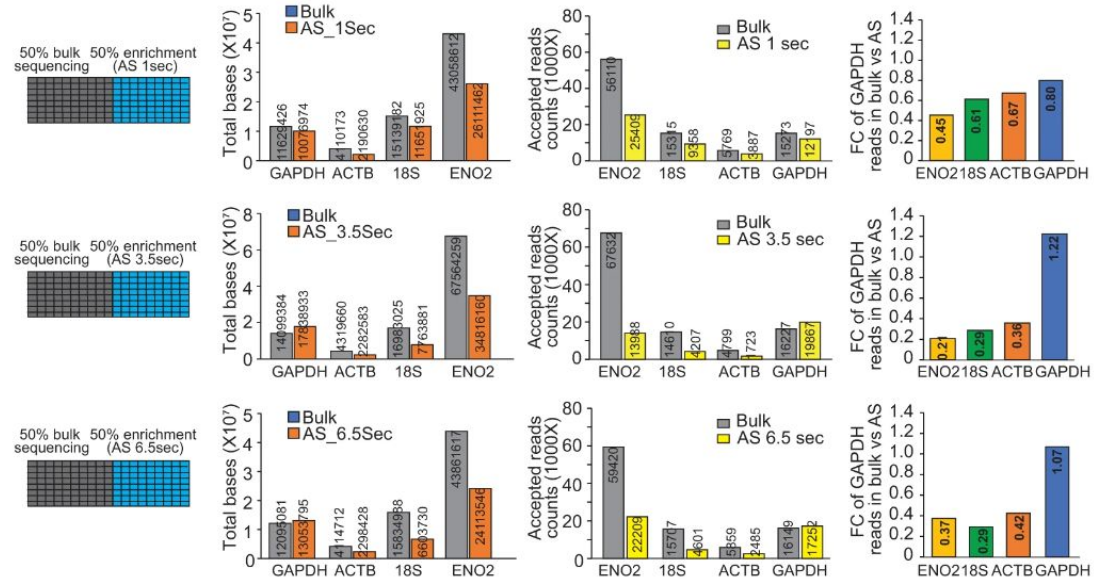


Image: Nanopore

Adaptive Sampling

Direct RNA sequencing (RNA004)

Nanopore
Community



Adaptive sampling performed on a pool of in vitro transcribed RNAs resulted in a net increase of 22-30% in the proportion of transcripts of interest in the population.

Wang, J., et al.. (2024). Direct RNA sequencing coupled with adaptive sampling enriches RNAs of interest in the transcriptome.
<https://doi.org/10.1038/s41467-023-44656-3>

Image: Nanopore

Section 2

Basecalling

Raw data formats and Basecalling

PacBio

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Official tool	ICS v13.3 (Revio)
	Base calling HiFi read generation with DeepConsensus Methylation calling Barcode demultiplexing BAM file generation

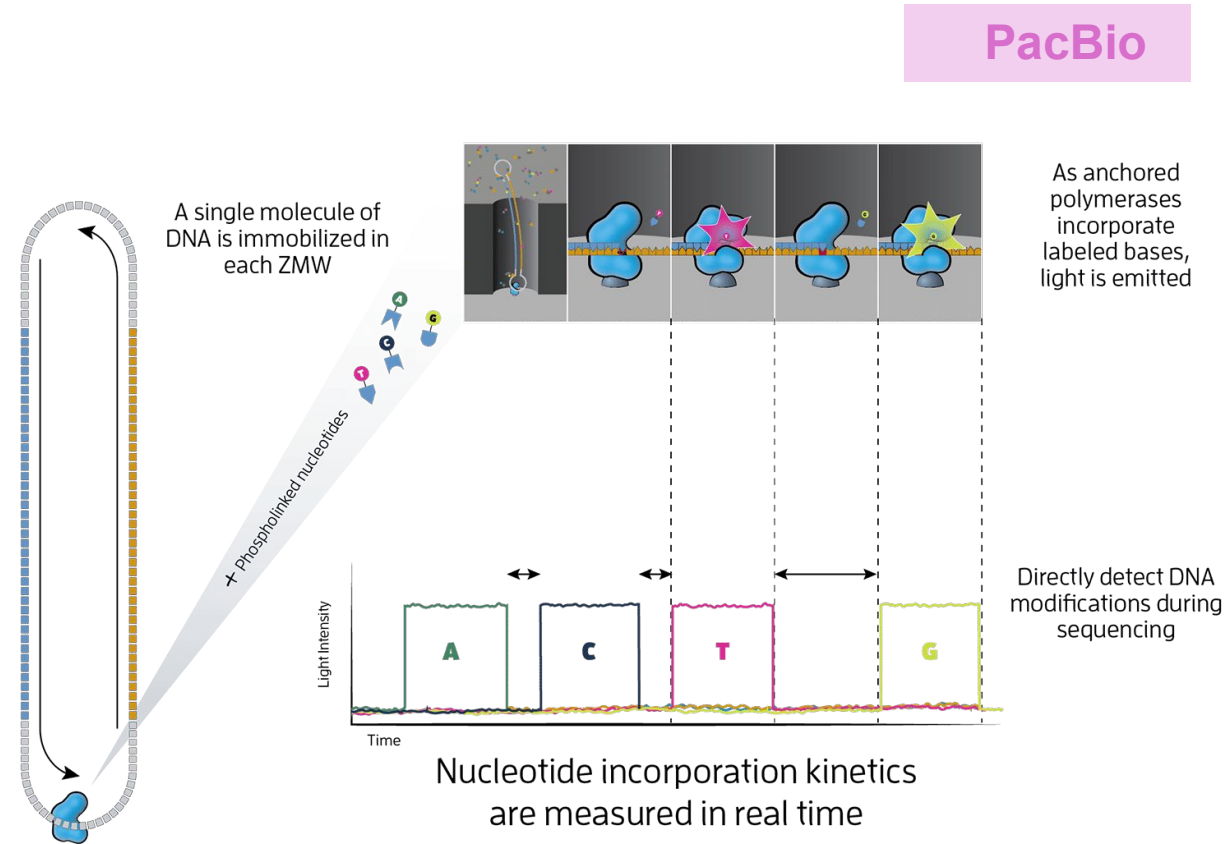


Image: Nanopore

PacBio

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Official tool	ICS v13.3 (Revio)
	Base calling HiFi read generation with DeepConsensus Methylation calling Barcode demultiplexing BAM file generation

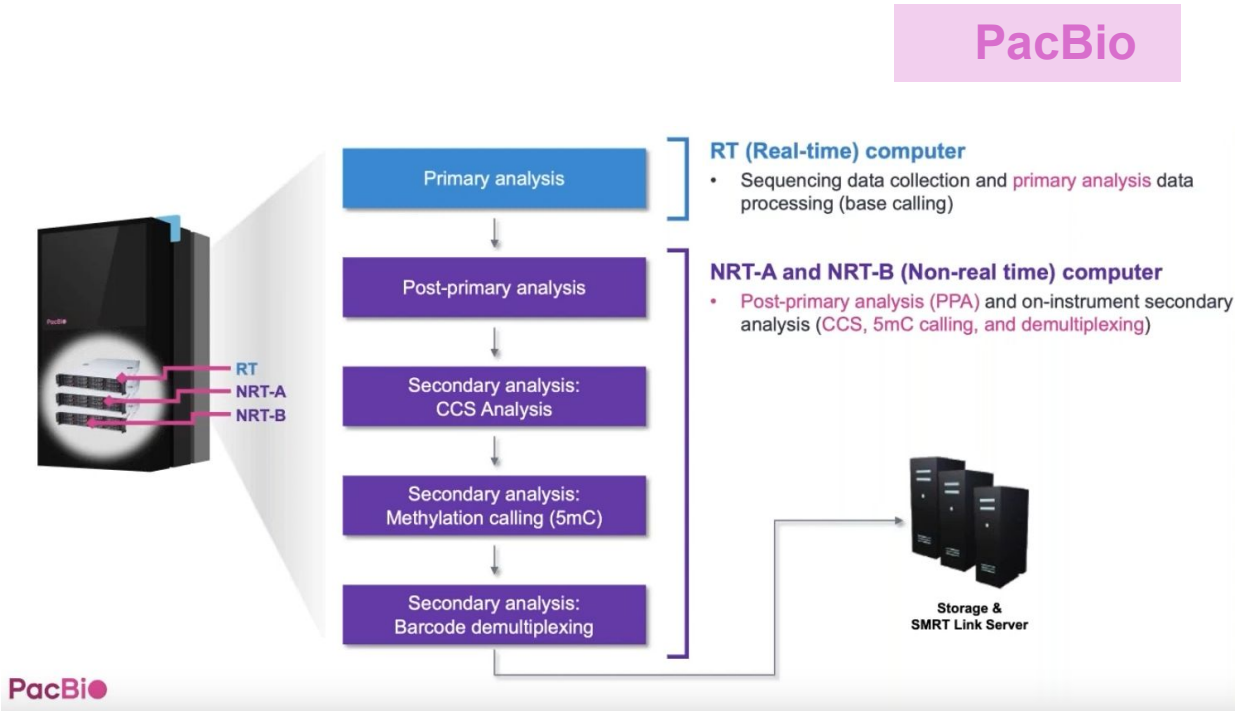


Image: Nanopore

Nanopore Basecalling

cDNA-PCR (R10.4.1)

Nanopore

Official tool	MinKNOW (dorado) Dorado standalone
Model	dna_r10.4.1_e8.2_400bps_fast@v5.2.0 dna_r10.4.1_e8.2_400bps_hac@v5.2.0 dna_r10.4.1_e8.2_400bps_sup@v5.2.0
Resource	PromethION Tower, P2I Professional/Gaming Laptop/Tower

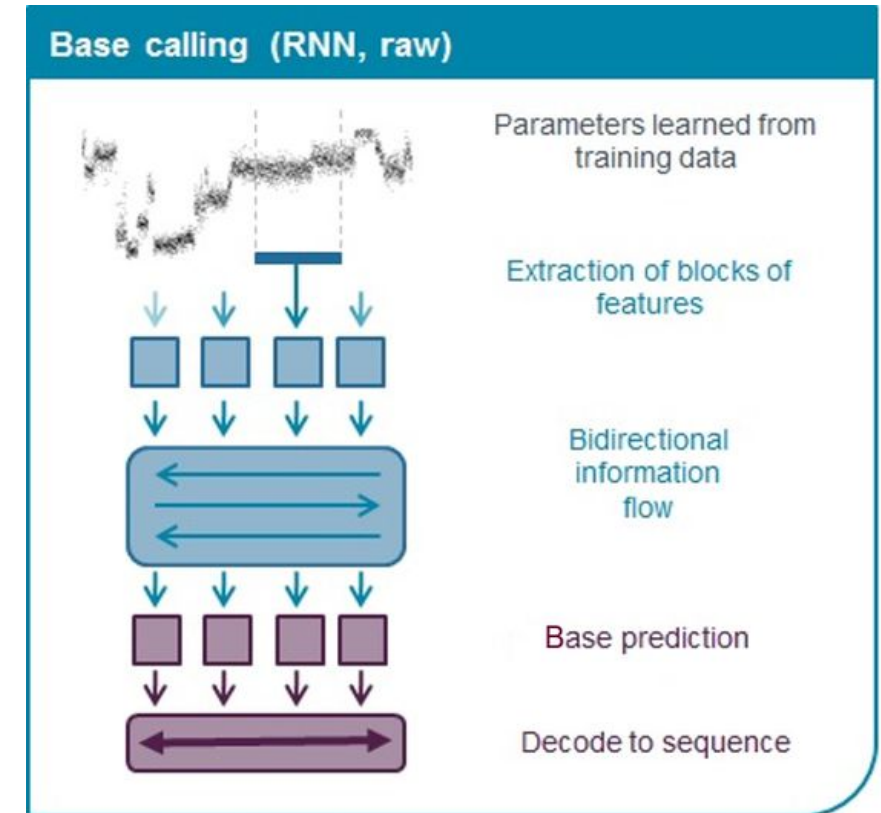


Image: Nanopore

Nanopore Basecalling

Direct RNA sequencing (RNA004)

Nanopore

Official tool	MinKNOW (dorado) Dorado standalone Remora
Model and mods	rna004_130bps_fast@v5.2.0 rna004_130bps_hac@v5.2.0 m5C m6A_DRACH inosine_m6A pseU rna004_130bps_sup@v5.2.0 m5C_2OmeC m6A_DRACH inosine_m6A_2OmeA pseU_2OmeU 2OmeG
Resource	PromethION Tower, P2I Professional/Gaming Laptop/Tower

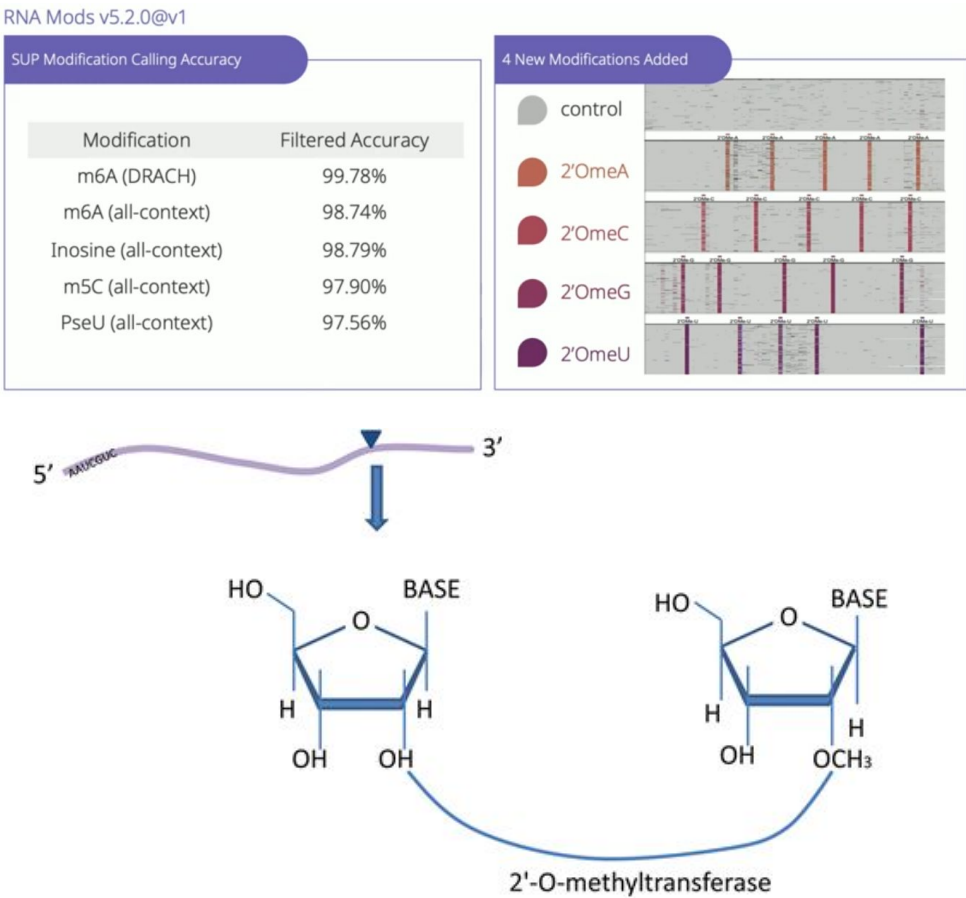


Image: Nanopore

Nanopore Modcalling

Direct RNA sequencing (RNA004)

Nanopore

Community tool

EpiNano (Liu et al, 2019)
m6A

Nanocompore (Leger, 2021)
De novo in comparison with IVT

m6anet (Hendra et al, 2022)
m6A

NanoRMS (Begik, 2022)
De novo, modification sites

m6ABasecaller (Cruciani, 2023)
m6A

and many more ...

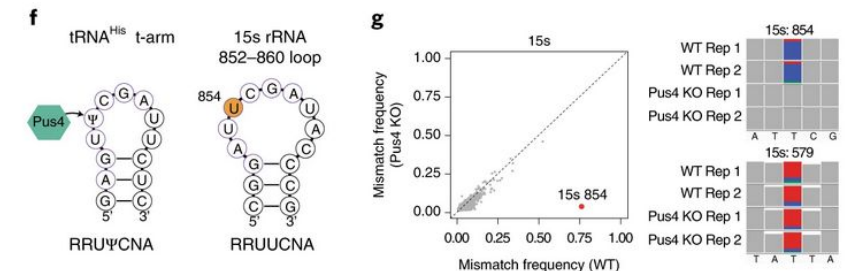
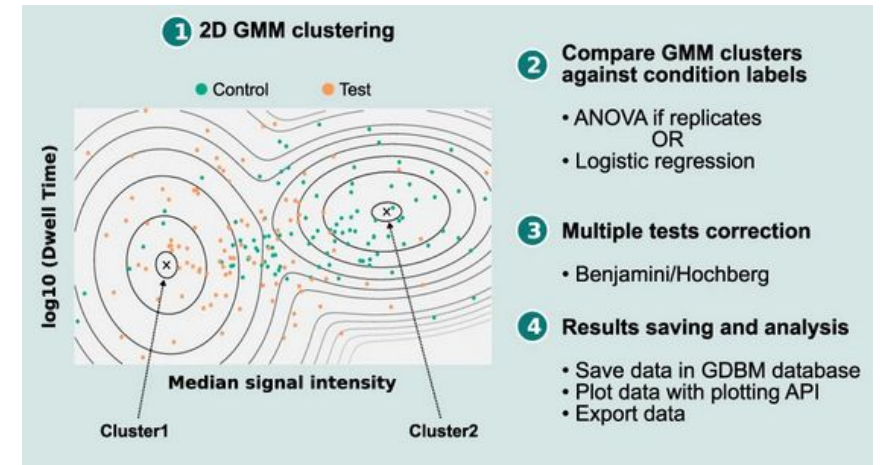


Image: Nanopore

Questions

1. What method kinnex was based on?
2. What improved throughput in Kinnex kit?
3. Why RT is performed in dRNA sequencing?
4. What mods supported by dorado?

Thank You!



For more information about the LongTREC Summer School:

<https://longtrec.eu>